Package: ctsfeatures (via r-universe)

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Type Package

Title Analyzing Categorical Time Series

Version 1.2.2

Description An implementation of several functions for feature extraction in categorical time series datasets. Specifically, some features related to marginal distributions and serial dependence patterns can be computed. These features can be used to feed clustering and classification algorithms for categorical time series, among others. The package also includes some interesting datasets containing biological sequences. Practitioners from a broad variety of fields could benefit from the general framework provided by 'ctsfeatures'.

License GPL-2

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LazyDataCompression xz

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Contents

binarization *Constructs the binarized time series associated with a given categorical time series*

Description

binarization constructs the binarized time series associated with a given categorical time series.

Usage

```
binarization(series)
```
Arguments

series An object of type tsibble (see R package tsibble), whose column named Value contains the values of the corresponding CTS. This column must be of class factor and its levels must be determined by the range of the CTS.

Details

Given a CTS of length T with range $V = \{1, 2, \ldots, r\}$, $\overline{X}_t = \{\overline{X}_1, \ldots, \overline{X}_T\}$, the function constructs the binarized time series, which is defined as $\overline{\bm{Y}}_t = \{\overline{\bm{Y}}_1,\ldots,\overline{\bm{Y}}_T\}$, with $\overline{\bm{Y}}_k = (\overline{Y}_{k,1},\ldots,\overline{Y}_{k,r})^\top$ such that $\overline{Y}_{k,i} = 1$ if $\overline{X}_k = i$ $(k = 1, ..., T, i = 1, ..., r)$. The binarized series is constructed in the form of a matrix whose rows represent time observations and whose columns represent the categories in the original series

Value

The binarized time series.

Author(s)

Ángel López-Oriona, José A. Vilar

References

López-Oriona Á, Vilar JA, D'Urso P (2023). "Hard and soft clustering of categorical time series based on two novel distances with an application to biological sequences." *Information Sciences*, 624, 467–492.

Examples

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
binarized_series <- binarization(sequence_1) # Constructing the binarized
# time series for the first CTS in dataset GeneticSequences
```
calculate_features *Computes several features associated with a categorical time series*

Description

calculate_features computes several features associated with a categorical time series or between a categorical and a real-valued time series

Usage

```
calculate_features(series, n_series = NULL, lag = 1, type = NULL)
```
Arguments

Details

Assume we have a CTS of length T with range $V = \{1, 2, ..., r\}$, $\overline{X}_t = \{\overline{X}_1, ..., \overline{X}_T\}$, with \hat{p}_i being the natural estimate of the marginal probability of the *i*th category, and $\hat{p}_{i,j}(l)$ being the natural estimate of the joint probability for categories i and j at lag l, $i, j = 1, \ldots, r$. Assume also that we have a real-valued time series of length $T, \overline{Z}_t = \{\overline{Z}_1, \ldots, \overline{Z}_T\}$. The function computes the following quantities depending on the argument type:

- If type=gini_index, the function computes the estimated gini index, $\hat{g} = \frac{r}{r-1}(1-\sum_{i=1}^r \hat{p}_i^2)$.
- If type=entropy, the function computes the estimated entropy, $\hat{e} = \frac{-1}{\ln(r)} \sum_{i=1}^r \hat{p}_i \ln \hat{p}_i$.
- If type=chebycheff_dispersion, the function computes the estimated chebycheff dispersion, $\hat{c} = \frac{r}{r-1} (1 - \max_i \hat{p}_i).$
- If type=gk_tau, the function computes the estimated Goodman and Kruskal's tau, $\hat{\tau}(l) = \sum_{i,j=1}^r \frac{\hat{p}_{ij}(l)^2}{\hat{p}_i} \sum_{i=1}^r \hat{p}_i^2$ $\frac{\widehat{p}_j^{(t)-}}{\widehat{p}_j}-\sum_{i=1}^r \widehat{p}_i^2$ $\frac{p_j}{1-\sum_{i=1}^r \widehat{p}_i^2}.$
- If type=gk_lambda, the function computes the estimated Goodman and Kruskal's lambda, $\widehat{\lambda}(l) = \frac{\sum_{j=1}^{r} \max_{i} \widehat{p}_{ij}(l)-\max_{i} \widehat{p}_{i}}{1-\max_{i} \widehat{p}_{i}}$ $\frac{\max_i p_{ij}(i) - \max_i p_i}{1 - \max_i \widehat{p}_i}.$
- If type=uncertainty_coefficient, the function computes the estimated uncertainty coefficient, $\widehat{u}(l) = -\frac{\sum_{i,j=1}^{r} \widehat{p}_{ij}(l) \ln \left(\frac{\widehat{p}_{ij}(l)}{\widehat{p}_{i} \widehat{p}_{j}} \right)}{\sum_{i=1}^{r} \widehat{p}_{i} \ln \widehat{p}_{i}}$ $\frac{\sum_{i=1}^r \widehat{p}_i \ln \widehat{p}_i}{\sum_{i=1}^r \widehat{p}_i \ln \widehat{p}_i}$.
- If type=pearson_measure, the function computes the estimated Pearson measure, $\hat{X}_T^2(l) =$ $T\sum_{i,j=1}^r \frac{(\widehat p_{ij}(l)-\widehat p_i\widehat p_j)^2}{\widehat p_i\widehat p_j}$ $\frac{\widehat{p}_i - p_i p_j}{\widehat{p}_i \widehat{p}_j}$.
- If type=phi2_measure, the function computes the estimated Phi2 measure, $\widehat{\Phi}^2(l) = \frac{\widehat{X}_T^2(l)}{T}$.
- If type=sakoda_measure, the function computes the estimated Sakoda measure, $\hat{p}^*(l) = \sqrt{\frac{2}{l} \sum_{k=1}^{l} p_k(l)}$ $\sqrt{\frac{r\widehat{\Phi}^2(l)}{(r-1)(1+\widehat{\Phi}^2(l))}}$.
- If type=cramers_vi, the function computes the estimated Cramer's vi, $\hat{v}(l) = \sqrt{\frac{1}{r-1} \sum_{i,j=1}^r \frac{(\hat{p}_{ij}(l) \hat{p}_i \hat{p}_j)^2}{\hat{p}_i \hat{p}_j}}$ $\frac{(p-p_ip_j)^2}{\widehat{p}_i\widehat{p}_j}$.
- If type=cohens_kappa, the function computes the estimated Cohen's kappa, $\hat{\kappa}(l) = \frac{\sum_{j=1}^{r}(\hat{p}_{jj}(l)-\hat{p}_j^2)}{1-\sum_{i=1}^{r}\hat{p}_i^2}$ $\frac{1}{1-\sum_{i=1}^r \hat{p}_i^2}$.
- If type=total_correlation, the function computes the the estimated sum $\widehat{\Psi}(l) = \frac{1}{r^2} \sum_{i,j=1}^r \widehat{\psi}_{ij}(l)^2$, where $\widehat{\psi}_{ij}(l)$ is the estimated correlation $\widehat{Corr}(Y_{t,i}, Y_{t-l,j}), i, j = 1, \ldots, r$, being $\overline{Y}_t = \widehat{Y_t}$ $\{\overline{Y}_1,\ldots,\overline{Y}_T\}$, with $\overline{Y}_k = (\overline{Y}_{k,1},\ldots,\overline{Y}_{k,r})^\top$, the binarized time series of \overline{X}_t .
- If type=spectral_envelope, the function computes the estimated spectral envelope.
- If type=total_mixed_correlation_1, the function computes the estimated total mixed lcorrelation given by

$$
\widehat{\Psi}_1(l) = \frac{1}{r} \sum_{i=1}^r \widehat{\psi}_i(l)^2,
$$

where $\widehat{\psi}_i(l) = \widehat{Corr}(Y_{t,i}, Z_{t-l})$, being $\overline{Y}_t = {\overline{Y}_1, \ldots, \overline{Y}_T}$, with $\overline{Y}_k = (\overline{Y}_{k,1}, \ldots, \overline{Y}_{k,r})^\top$, the binarized time series of \overline{X}_t .

• If type=total_mixed_correlation_2, the function computes the estimated total mixed qcorrelation given by

$$
\widehat{\Psi}_2(l) = \frac{1}{r} \sum_{i=1}^r \int_0^1 \widehat{\psi}_i^{\rho}(l)^2 d\rho,
$$

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where $\hat{\psi}_i^{\rho}(l) = \widehat{Corr}(Y_{t,i}, I(Z_{t-l} \leq q_{Z_t}(\rho)))$, being $\overline{Y}_t = {\overline{Y}_1, \dots, \overline{Y}_T}$, with $\overline{Y}_k =$ $(\overline{Y}_{k,1},\ldots,\overline{Y}_{k,r})^{\top}$, the binarized time series of \overline{X}_t , $\rho \in (0,1)$ a probability level, $I(\cdot)$ the indicator function and q_{Z_t} the quantile function of the corresponding real-valued process.

Value

The corresponding feature.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH, Göb R (2008). "Measuring serial dependence in categorical time series." *AStA Advances in Statistical Analysis*, 92, 71–89.

Examples

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
uc <- calculate_features(series = sequence_1, type = 'uncertainty_coefficient' )
# Computing the uncertainty coefficient
# for the first series in dataset GeneticSequences
se <- calculate_features(series = sequence_1, type = 'spectral_envelope' )
# Computing the spectral envelope
# for the first series in dataset GeneticSequences
```
calculate_motifs *Computes the relative frequency of motifs in a categorical time series*

Description

calculate_motifs computes the motifs of a categorical time series

Usage

calculate_motifs(series, motif_length)

Arguments

Details

Given a CTS of length T with range $V = \{1, 2, ..., r\}$, $\overline{X}_t = \{\overline{X}_1, ..., \overline{X}_T\}$, and a motif length L, the function returns an array of r^L elements, with the element in the position (i_1, i_2, \ldots, i_r) being the relative frequency of the motif " $i_1i_2 \cdots i_r$ " in the corresponding time series.

Value

Returns an array with the relative frequency of motifs in a categorical time series.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Lonardi JLEKS, Patel P (2002). "Finding motifs in time series." In *Proc. of the 2nd Workshop on Temporal Data Mining*, 53–68.

Examples

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
calculate_motifs(sequence_1, motif_length = 3)
# Computing the relative frequencies of motifs of length 3 for the first
# series in dataset GeneticSequences
```
calculate_subfeatures *Computes several subfeatures associated with a categorical time series*

Description

calculate_features computes several subfeatures associated with a categorical time series or between a categorical and a real-valued time series

Usage

```
calculate_subfeatures(series, n_series, lag = 1, type = NULL)
```
Arguments

Details

Assume we have a CTS of length T with range $V = \{1, 2, ..., r\}$, $\overline{X}_t = \{\overline{X}_1, ..., \overline{X}_T\}$, with \hat{p}_i being the natural estimate of the marginal probability of the *i*th category, and $\hat{p}_{ij}(l)$ being the natural estimate of the joint probability for categories i and j at lag l, $i, j = 1, \ldots, r$. Assume also that we have a real-valued time series of length $T, Z_t = \{Z_1, \ldots, Z_T\}$. The function computes the following subfeatures depending on the argument type:

- If type=entropy, the function computes the subfeatures associated with the estimated entropy, $\widehat{p}_i \ln(\widehat{p}_i), i = 1, 2, \ldots, r$.
- If type=gk_tau, the function computes the subfeatures associated with the estimated Goodman and Kruskal's tau, $\frac{\widehat{p}_{ij}(l)^2}{\widehat{x}_{ij}}$ $\frac{i^{(t)}}{\widehat{p}_j}, i, j = 1, 2, \ldots, r.$
- If type=gk_lambda, the function computes the subfeatures associated with the estimated Goodman and Kruskal's lambda, $\max_i \widehat{p}_{i,j}(l), i = 1, 2, \ldots, r$.
- If type=uncertainty_coefficient, the function computes the subfeatures associated with the estimated uncertainty coefficient, $\hat{p}_{ij}(l) \ln \left(\frac{\hat{p}_{ij}(l)}{\hat{p}_i \hat{p}_j} \right)$ $\widehat{p}_i\widehat{p}_j$ $\big), i, j = 1, 2, \ldots, r.$
- If type=pearson_measure, the function computes the subfeatures associated with the estimated Pearson measure, $\frac{(\widehat{p}_{ij}(l)-\widehat{p}_i\widehat{p}_j)^2}{\widehat{p}_i\widehat{p}_j}$ $\frac{i(j-p_ip_j)}{\widehat{p}_i\widehat{p}_j}, i, j = 1, 2, \ldots, r.$
- If type=phi2_measure, the function computes the subfeatures associated with the estimated Phi2 measure, $\frac{(\widehat{p}_{ij}(l)-\widehat{p}_i\widehat{p}_j)^2}{\widehat{p}_i\widehat{p}_j}$ $\frac{i(j-p_ip_j)}{\widehat{p}_i\widehat{p}_j}, i, j = 1, 2, \ldots, r.$
- If type=sakoda_measure, the function computes the subfeatures associated with the estimated Sakoda measure, $\frac{(\widehat{p}_{ij}(l)-\widehat{p}_i\widehat{p}_j)^2}{\widehat{p}_i\widehat{p}_j}$ $\frac{i(j-p_ip_j)}{\widehat{p}_i\widehat{p}_j}, i, j = 1, 2, \ldots, r.$
- If type=cramers_vi, the function computes the subfeatures associated with the estimated Cramer's vi, $\frac{(\widehat{p}_{ij}(l)-\widehat{p}_i\widehat{p}_j)^2}{\widehat{p}_i\widehat{p}_j}$ $\frac{i(j-p_ip_j)}{\widehat{p}_i\widehat{p}_j}, i, j = 1, 2, \ldots, r.$
- If type=cohens_kappa, the function computes the subfeatures associated with the estimated Cohen's kappa, $\widehat{p}_{ii}(l) - \widehat{p}_i^2$, $i = 1, 2, \ldots, r$.
- If type=total_correlation, the function computes the subfeatures associated with the total correlation, $\widehat{\psi}_{ij}(l), i, j = 1, 2, \ldots, r$ (see type='total_mixed_cor' in the function calculate_features).
- If type=total_mixed_correlation_1, the function computes the subfeatures associated with the total mixed l-correlation, $\psi_i(l), i = 1, 2, \ldots, r$ (see type='total_mixed_correlation_1' in the function calculate_features).
- If type=total_mixed_correlation_2, the function computes the subfeatures associated with the total mixed q-correlation, $\int_0^1 \widehat{\psi}_i^{\rho}(l)^2 d\rho, i = 1, 2, \ldots, r$ (see type='total_mixed_correlation_2' in the function calculate_features).

Value

The corresponding subfeature

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH, Göb R (2008). "Measuring serial dependence in categorical time series." *AStA Advances in Statistical Analysis*, 92, 71–89.

Examples

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
suc <- calculate_subfeatures(series = sequence_1, type = 'uncertainty_coefficient')
# Computing the subfeatures associated with the uncertainty coefficient
# for the first series in dataset GeneticSequences
scv <- calculate_subfeatures(series = sequence_1, type = 'cramers_vi' )
# Computing the subfeatures associated with the cramers vi
# for the first series in dataset GeneticSequences
```
conditional_probabilities

Computes the conditional probabilities of a categorical time series

Description

conditional_probabilities returns a matrix with the conditional probabilities of a categorical time series

Usage

```
conditional_probabilities(series, lag = 1)
```
Arguments

Details

Given a CTS of length T with range $V = \{1, 2, \ldots, r\}$, $\overline{X}_t = \{\overline{X}_1, \ldots, \overline{X}_T\}$, the function computes the matrix $\widehat{P}^{\vec{c}}(l) = (\widehat{p}^{\vec{c}}_{ij}(l))_{1 \leq i,j \leq r}$, with $\widehat{p}^{\vec{c}}_{ij}(l) = \frac{TN^{\vec{c}}_{ij}(l)}{(T-l)N_i}$, where N_i is the number of elements equal to i in the realization \overline{X}_t and $N_{ij}(l)$ is the number of pairs $(\overline{X}_t, \overline{X}_{t-l}) = (i, j)$ in the realization \overline{X}_t .

Value

A matrix with the conditional probabilities.

Author(s)

Ángel López-Oriona, José A. Vilar

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References

Weiß CH, Göb R (2008). "Measuring serial dependence in categorical time series." *AStA Advances in Statistical Analysis*, 92, 71–89.

Examples

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
matrix_cp <- conditional_probabilities(series = sequence_1) # Computing the matrix of
# joint probabilities for the first series in dataset GeneticSequences
```
GeneticSequences *GeneticSequences*

Description

Categorical time series (CTS) of DNA sequences from different viruses

Usage

data(GeneticSequences)

Format

A tsibble with four columns, which are:

Value The categorical values of the time series in the dataset.

Series Integer values indicating the considered time series (there are 32 time series in the dataset).

Time Integer values indicating the temporal indexes of the observations.

Class Integer values indicating the class of each time series.

Details

The column Value is the concatenation of 32 time series taking four categorical values (DNA bases). The column Class is formed by integers from 1 to 4, indicating that there are 4 different classes in the database. Each class is associated with a different family of viruses. For more information, see López-Oriona et al. (2023).

References

López-Oriona Á, Vilar JA, D'Urso P (2023). "Hard and soft clustering of categorical time series based on two novel distances with an application to biological sequences." *Information Sciences*, 624, 467–492.

joint_probabilities *Computes the joint probabilities of a categorical time series*

Description

joint_probabilities returns a matrix with the joint probabilities of a categorical time series

Usage

```
joint_probabilities(series, lag = 1)
```
Arguments

Details

Given a CTS of length T with range $V = \{1, 2, \ldots, r\}$, $\overline{X}_t = \{\overline{X}_1, \ldots, \overline{X}_T\}$, the function computes the matrix $\hat{P}(l) = (\hat{p}_{ij}(l))_{1 \leq i,j \leq r}$, with $\hat{p}_{ij}(l) = \frac{N_{ij}(l)}{T-l}$, where $N_{ij}(l)$ is the number of pairs $(\overline{X}_t, \overline{X}_{t-l}) = (i, j)$ in the realization \overline{X}_t .

Value

A matrix with the joint probabilities.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH, Göb R (2008). "Measuring serial dependence in categorical time series." *AStA Advances in Statistical Analysis*, 92, 71–89.

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
matrix_jp <- joint_probabilities(series = sequence_1) # Computing the matrix of
# joint probabilities for the first series in dataset GeneticSequences
```
marginal_probabilities

Computes the marginal probabilities of a categorical time series

Description

marginal_probabilities returns a vector with the marginal probabilities of a categorical time series

Usage

marginal_probabilities(series)

Arguments

series An object of type tsibble (see R package tsibble), whose column named Value contains the values of the corresponding CTS. This column must be of class factor and its levels must be determined by the range of the CTS.

Details

Given a CTS of length T with range $V = \{1, 2, \ldots, r\}$, $\overline{X}_t = \{\overline{X}_1, \ldots, \overline{X}_T\}$, the function computes the vector $\hat{\mathbf{p}} = (\hat{p}_1, \dots, \hat{p}_r)$, with $\hat{p}_i = \frac{N_i}{T}$, where N_i is the number of elements equal to i in the realization \overline{X}_t .

Value

A vector with the marginal probabilities.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH, Göb R (2008). "Measuring serial dependence in categorical time series." *AStA Advances in Statistical Analysis*, 92, 71–89.

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
vector_mp <- marginal_probabilities(series = sequence_1) # Computing the vector of
# marginal probabilities for the first series in dataset GeneticSequences
```


Description

plot_ccc constructs a control chart for the cycle lengths of a categorical series

Usage

```
plot_ccc(
  series,
  mu_t,
  lcl_t,
  ucl_t,
  plot = TRUE,
  title = "Control chart (cycles)",
  ...
)
```
Arguments

Details

Constructs a control chart of a CTS based on cycle lengths. The chart is based on the standardized statistic $T_t = T_t^{(L)} + T_t^{(U)}$, with $T_t^{(L)} = \min\left(0, \frac{C_t - \mu_t}{|LCL_t - \mu_t|}\right)$ and $T_t^{(U)} = \max\left(0, \frac{C_t - \mu_t}{|UCL_t - \mu_t|}\right)$, where Z_t expresses the length of a cycle ending with a specific category, μ_t denotes the mean of Z_t and LCL_t and UCL_t are lower and upper individual control limits, respectively. Note that an out-of-control alarm is signalled if $T_t < -1$ or $T_t > 1$.

Value

If plot = TRUE (default), represents the control chart for the cycle lengths. Otherwise, the function returns a matrix with the values of the standardized statistic for each time t

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Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH (2008). "Visual analysis of categorical time series." *Statistical Methodology*, 5(1), 56–71.

Examples

```
sequence_1 <- SyntheticData1[which(SyntheticData1$Series==1),]
cycle\_cc \leftarrow plot\_ccc(series = sequence_1, mu_t = c(1, 1.5, 1),lcl_t = rep(10, 600), ucl_t = rep(10, 600)) # Representing
# a control chart for the cycle lengths
cycle\_cc \leftarrow plot\_ccc(series = sequence_1, mu_t = c(1, 1.5, 1),lcl_t = rep(10, 600), ucl_t = rep(10, 600), plot = FALSE) # Computing the
# corresponding standardized statistic
```
plot_cohen *Constructs a serial dependence plot based on Cohen's kappa*

Description

plot_cohen constructs a serial dependence plot of a categorical time series based on Cohen's kappa

Usage

```
plot_cohen(
  series,
 max\_lag = 10,
 alpha = 0.05,
 plot = TRUE,
  title = "Serial dependence plot",
 bar_wwidth = 0.12,
  ...
\lambda
```
Arguments

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Details

Constructs a serial dependence plot based on Cohens's kappa, $\hat{\kappa}(l)$, for several lags. A dashed lined is incorporated indicating the critical value of the test based on the following asymptotic approximation (under the i.i.d. assumption):

$$
\sqrt{\frac{T}{V(\widehat{\boldsymbol{p}})}}\left(\widehat{\kappa}(l)+\frac{1}{T}\right) \sim N(0,1),
$$

where T is the series length, $\hat{p} = (\hat{p}_1, \dots, \hat{p}_r)$ is the vector of estimated marginal probabilities for the *r* categories of the series and $V(\hat{p}) = 1 - \frac{1+2\sum_{i=1}^{r} \hat{p}_i^3 - 3\sum_{i=1}^{r} \hat{p}_i^2}{(1-\sum_{i=1}^{r} \hat{p}_i^2)^2}$.

Value

If plot = TRUE (default), returns the serial dependence plot based on Cohens's kappa. Otherwise, the function returns a list with the values of Cohens's kappa, the critical value and the corresponding p-values.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH (2011). "Empirical measures of signed serial dependence in categorical time series." *Journal of Statistical Computation and Simulation*, 81(4), 411–429.

Examples

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
plot_ck <- plot_cohen(series = sequence_1, max_lag = 3) # Representing
# the serial dependence plot
list_ck <- plot_cohen(series = sequence_1, max_lag = 3, plot = FALSE) # Obtaining
# the values of Cohens's kappa, the critical value and the p-values
```
plot_cramer *Constructs a serial dependence plot based on Cramer's vi*

Description

plot_cramer constructs a serial dependence plot of a categorical time series based on Cramer's vi

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Usage

```
plot_cramer(
  series,
  max\_lag = 10,
  alpha = 0.05,
 plot = TRUE,
  title = "Serial dependence plot",
 bar_wwidth = 0.12,
  ...
)
```
Arguments

Details

Constructs a serial dependence plot based on Cramer's vi, $\hat{v}(l)$, for several lags. A dashed lined is incorporated indicating the critical value of the test based on the following asymptotic approximation (under the i.i.d. assumption):

$$
T(r-1)\hat{v}(l)^{2} \sim \chi^{2}_{(r-1)^{2}},
$$

where T is the series length and r is the number of categories in the time series.

Value

If plot = TRUE (default), returns the serial dependence plot based on Cramer's vi. Otherwise, the function returns a list with the values of Cramer's vi, the critical value and the corresponding pvalues.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH (2013). "Serial dependence of NDARMA processes." *Computational Statistics and Data Analysis*, 68, 213–238.

Examples

```
sequence_1 <- SyntheticData1[which(SyntheticData1$Series==1),]
plot_cv <- plot_cramer(series = sequence_1, max_lag = 3) # Representing
# the serial dependence plot
list_cv <- plot_cramer(series = sequence_1, max_lag = 3, plot = FALSE) # Obtaining
# the values of Cramer's vi, the critical value and the p-values
```
plot_cts *Constructs a categorical time series plot*

Description

plot_cts constructs a categorical time series plot

Usage

```
plot_cts(series, title = "Time series plot")
```
Arguments

Details

Constructs a categorial time series plot for a given CTS.

Value

The categorical time series plot.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH (2018). *An introduction to discrete-valued time series*. John Wiley and Sons.

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
time_series_plot <- plot_cts(series = sequence_1) # Constructs a categorical
# time series plot for the first 50 observations of the first time series in
# dataset GeneticSequences
```


Description

plot_ifsct constructs the IFS circle transformation of a categorical time series.

Usage

```
plot_ifsct(series, alpha, beta, title = "IFS circle transformation", ...)
```
Arguments

Details

Constructs the IFS circle transformation for a given CTS, which is useful to identify cycles of arbitrary length.

Value

The IFS circle transformation.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH (2008). "Visual analysis of categorical time series." *Statistical Methodology*, 5(1), 56–71.

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
ct <- plot_ifsct(sequence_1, alpha = 0.1, beta = 0.1) # Constructing the IFS circle transformation
# for the first CTS in dataset GeneticSequences
```


Description

plot_mcc constructs a control chart for the marginal distribution of a categorical series

Usage

```
plot_mcc(
  series,
 c,
 sigma,
 lambda = 0.99,
 k = 3.3,min\_max = FALSE,plot = TRUE,
 title = "Control chart (marginal)",
  ...
)
```
Arguments

plot_ph the contract of the co

Details

Constructs a control chart of a CTS with range $V = \{1, \ldots, r\}$ based on the marginal distribution. The chart relies on the standardized statistic $T_{t,i} = \frac{\hat{\pi}_{t,i}^{(\lambda)} - p_i}{k \cdot \hat{\sigma}_{t,i}}$ $\frac{\sum_{t,i}^{(x)} - p_i}{k \cdot \sigma_{t,i}}$, where the $\hat{\pi}^{\left(\lambda\right)}_{t,i}, i = 1, \ldots, r$, are the components of the EWMA estimator of the marginal distribution, p_i is the marginal probability of category *i*, $\sigma_{t,i}$ is the variance of $\hat{\pi}_{t,i}^{(\lambda)}$ and *k* is a constant set by the user. If min_max = FALSE, then only the statistics $T_t^{\min} = \min_{i \in \mathcal{V}} T_{t,i}$ and $T_t^{\max} = \max_{i \in \mathcal{V}} T_{t,i}$ are plotted. An out-of-control alarm is signalled if the statistics are below -1 or above 1.

Value

If plot = TRUE (default), represents the control chart for the marginal distribution. Otherwise, the function returns a matrix with the values of the standardized statistics for each time t

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH (2008). "Visual analysis of categorical time series." *Statistical Methodology*, 5(1), 56–71.

Examples

```
sequence_1 <- SyntheticData1[which(SyntheticData1$Series==1),]
cycle\_cc \leftarrow plot\_ccc(series = sequence_1, mu_t = c(1, 1.5, 1),lcl_t = rep(10, 600), ucl_t = rep(10, 600))cycle_md <- plot_mcc(series = sequence_1, c = c(0.3, 0.3, 0.4),
sigma = matrix(rep(c(1, 1, 1), 600), nrow = 600)) # Representing
# a control chart for the marginal distribution
cycle_md <- plot_mcc(series = sequence_1, c = c(0.3, 0.3, 0.4),
sigma = matrix(rep(c(1, 1, 1), 600), nrow = 600), plot = FALSE) # Computing the
# corresponding standardized statistic
```
plot_ph *Constructs the pattern histogram associated with a given category of a categorical time series*

Description

plot_ph constructs the pattern histogram associated with a given category of a categorical time series.

Usage

```
plot_ph(
  series,
  category,
 plot = TRUE,
 title = paste0("Pattern histogram (", category, ")"),
  ...
)
```
Arguments

Details

Constructs the pattern histogram for a specific category of a CTS. This graph represents the frequencies of the cycles for the corresponding category according to their length.

Value

The pattern histogram.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Weiß CH (2008). "Visual analysis of categorical time series." *Statistical Methodology*, 5(1), 56–71.

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
ph <- plot_ph(sequence_1,
category = 'a') # Constructing the pattern histogram
# for the first CTS in dataset GeneticSequences concerning the category 'a'
cycle_lengths <- plot_ph(sequence_1,
category = 'a', plot = FALSE) # Obtaining the frequencies of cycle lengths
```


Description

plot_reg constructs the rate evolution graph proposed by Ribler (1997).

Usage

```
plot_reg(
  series,
  title = "Rate evolution graph",
  linear_fit = FALSE,
  cat_res = NULL,
  ...
)
```
Arguments

Details

Given a CTS of length T with range $V = \{1, 2, ..., r\}$, $X_t = \{X_1, ..., X_T\}$, and the corresponding binarized time series, $\overline{Y}_t = \{\overline{Y}_1, \ldots, \overline{Y}_T\}$, the function constructs the rate evolution graph. Specifically, consider the series of cumulated sums given by $\overline{C}_t = \{\overline{C}_1, \ldots, \overline{C}_T\}$, with $\overline{C}_k = \sum_{s=1}^k \overline{Y}_s$, $k = 1, \ldots, T$. The rate evolution graph displays a standard time series plot for each one of the components of C_t simultaneously in one graph.

Value

The rate evolution graph.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Ribler RL (1997). *Visualizing categorical time series data with applications to computer and communications network traces*. Ph.D. thesis, Virginia Polytechnic Institute and State University.

Examples

sequence_1 <- GeneticSequences[which(GeneticSequences\$Series==1),] reg <- plot_reg(sequence_1) # Constructing the rate # evolution graph for the first time series in dataset GeneticSequences

plot_se *Represents the spectral envelope of a categorical time series*

Description

plot_se represents the spectral envelope of a categorical time series

Usage

plot_se(series)

Arguments

series An object of type tsibble (see R package tsibble), whose column named Value contains the values of the corresponding CTS. This column must be of class factor and its levels must be determined by the range of the CTS.

Details

The function represents the spectral envelope of a categorical time series

Value

Returns returns a plot of the spectral envelope.

Author(s)

Ángel López-Oriona, José A. Vilar

References

Stoffer DS, Tyler DE, McDougall AJ (1993). "Spectral analysis for categorical time series: Scaling and the spectral envelope." *Biometrika*, 80(3), 611–622.

```
sequence_1 <- GeneticSequences[which(GeneticSequences$Series==1),]
plot_se(sequence_1)
# Representing the spectral envelope for the first series in dataset
# GeneticSequences
```


ProteinSequences *ProteinSequences*

Description

Categorical time series (CTS) of protein sequences from different species

Usage

```
data(ProteinSequences)
```
Format

A tsibble with four columns, which are:

Value The categorical values of the time series in the dataset.

Series Integer values indicating the considered time series (there are 40 time series in the dataset).

Time Integer values indicating the temporal indexes of the observations.

Class Integer values indicating the class of each time series.

Details

The column Value is the concatenation of 40 time series taking four categorical values (aminoacids). The column Class is formed by integers from 1 to 4, indicating that there are 4 different classes in the database. Each class is associated with a different family of viruses. For more information, see López-Oriona et al. (2023).

References

López-Oriona Á, Vilar JA, D'Urso P (2023). "Hard and soft clustering of categorical time series based on two novel distances with an application to biological sequences." *Information Sciences*, 624, 467–492.

SleepStages *SleepStages*

Description

Categorical time series (CTS) of sleep stages from different subjects

Usage

data(SleepStages)

Format

A tsibble with four columns, which are:

Value The categorical values of the time series in the dataset.

Series Integer values indicating the considered time series (there are 62 time series in the dataset).

Time Integer values indicating the temporal indexes of the observations.

Class Integer values indicating the class of each time series.

Details

The column Value is the concatenation of 62 time series taking six categorical values (sleep stages). The column Class is formed by the integers 1 and 2 indicating that there are 2 different classes in the database. Each class is associated with a sleep disorder (class 1 refers to nocturnal frontal lobe epilepsy, while class refers 2 to REM behavior disorder). For more information, see López-Oriona et al. (2023).

References

López-Oriona Á, Vilar JA, D'Urso P (2023). "Hard and soft clustering of categorical time series based on two novel distances with an application to biological sequences." *Information Sciences*, 624, 467–492.

SyntheticData1 *SyntheticData1*

Description

Synthetic dataset containing 80 CTS generated from four different generating processes.

Usage

```
data(SyntheticData1)
```
Format

A tsibble with four columns, which are:

Value The categorical values of the time series in the dataset.

Series Integer values indicating the considered time series (there are 80 time series in the dataset).

Time Integer values indicating the temporal indexes of the observations.

Class Integer values indicating the class of each time series.

@details The column Value is the concatenation of 80 time series of length 600 taking three categorical values. Series 1-20, 21-40, 41-60 and 61-80 were generated from Markov Chains with different matrices of transition probabilities (see Scenario 1 in López-Oriona et al. (2023)). Therefore, there are 4 different classes in the dataset.

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References

López-Oriona Á, Vilar JA, D'Urso P (2023). "Hard and soft clustering of categorical time series based on two novel distances with an application to biological sequences." *Information Sciences*, 624, 467–492.

SyntheticData2 *SyntheticData2*

Description

Synthetic dataset containing 80 CTS generated from four different generating processes.

Usage

data(SyntheticData2)

Format

A tsibble with four columns, which are:

Value The categorical values of the time series in the dataset.

Series Integer values indicating the considered time series (there are 80 time series in the dataset).

Time Integer values indicating the temporal indexes of the observations.

Class Integer values indicating the class of each time series.

@details The column Value is the concatenation of 80 time series of length 600 taking three categorical values. Series 1-20, 21-40, 41-60 and 61-80 were generated from Hidden Markov Models with different matrices of transition and emission probabilities (see Scenario 2 in López-Oriona et al. (2023)). Therefore, there are 4 different classes in the dataset.

References

López-Oriona Á, Vilar JA, D'Urso P (2023). "Hard and soft clustering of categorical time series based on two novel distances with an application to biological sequences." *Information Sciences*, 624, 467–492.

SyntheticData3 *SyntheticData3*

Description

Synthetic dataset containing 80 CTS generated from four different generating processes.

Usage

data(SyntheticData3)

Format

A tsibble with four columns, which are:

Value The categorical values of the time series in the dataset.

Series Integer values indicating the considered time series (there are 80 time series in the dataset).

Time Integer values indicating the temporal indexes of the observations.

Class Integer values indicating the class of each time series.

@details The column Value is the concatenation of 80 time series of length 600 taking three categorical values. Series 1-20, 21-40, 41-60 and 61-80 were generated from NDARMA processes with different orders and vectors of coefficients (see Scenario 3 in López-Oriona et al. (2023)). Therefore, there are 4 different classes in the dataset.

References

López-Oriona Á, Vilar JA, D'Urso P (2023). "Hard and soft clustering of categorical time series based on two novel distances with an application to biological sequences." *Information Sciences*, 624, 467–492.

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